## 2008*Phytophthora ramorum* Survey for the Northern Sierra Nevada California Department of Forestry and Fire Protection Submitted by Donald R. Owen December 2008

### Summary

A stream-based detection survey was conducted in Butte, Yuba, Nevada, Placer, and El Dorado Counties in northern California for *Phytophthora ramorum*, the cause of sudden oak death. A total of 31 watercourse sites were surveyed during May and June using rhododendron leaves as bait for *Phytophthora* spp. This produced 118 sets of baits that were cultured for the presence of *P. ramorum*. The pathogen was not detected in any of the samples. Other *Phytophthora spp.* were recovered but not identified.

## SUDDEN OAK DEATH / P. ramorum SURVEY

The quarantined disease organism *P. ramorum* is not established in the Sierra Nevada of California, although hosts and putatively suitable habitat occur there. Risk analyses (Meentemeyer et al. 2004\*) identified the foothills of the northern Sierra Nevada, from El Dorado County north to Butte County, as having the most suitable habitat for establishment of *P. ramorum* within the mountain range. This is the 5th year that some portion of this area has been surveyed for *P. ramorum*. This year's objectives were to 1) use rhododendron baits to recover *Phytophthora spp.* from the principal watercourses draining these habitats, 2) to expand and further refine the watercourse sampling sites within the area, and 3) conduct follow-up surveys if *P. ramorum* was recovered.

The project was planned and coordinated by CDF Entomologist Don Owen. Retired CDF Forester Don MacKenzie was the principal surveyor. Surveys were conducted from early May through late June 2008. Kamyar Aram (Dave Rizzo's Lab, UC Davis) provided the baits and conducted the diagnostics for watercourse samples. Surveys were conducted on private land, portions of the Plumas, Tahoe and Eldorado National Forests, and several State Recreation Areas. Personnel from the Yuba River Ranger District provided transportation across New Bullard's Bar Reservoir and assisted with surveys at remote watercourse sites there.

## Procedures

In 2006, Douglas Shoemaker of the Center of Applied GIS (under the direction of Ross Meentemeyer), University of North Carolina, provided 1:100,000 scale maps delineating

<sup>\*</sup> Meentemeyer, R, D Rizzo, W Mark, and E Lotz. 2004. Mapping the risk of establishment and spread of sudden oak death in California. For Ecol and Mgmt. 200: 195–214.

areas of moderate to high risk for the establishment of *P. ramorum* in the northern Sierra Nevada. These were overlaid with standard 1:100,000 USGS maps to identify access roads and watercourses that traverse high-risk habitats. Locations of potential watercourse-sampling sites were identified and plotted on 1:24,000 scale topographic maps and 1:12,000 orthophotos for field use. Following a reconnaissance of potential sites, a total of 23 watercourses were sampled in 2006 in Butte, Yuba, Nevada, Placer, and El Dorado Counties. A number of sites were moved or added in 2007 and 2008, resulting in 28 and 31 watercourse sites, respectively, being sampled during these years. The principal criteria for site selection are the amount of high risk habitat that exists upstream of a site and access.

Watercourses were sampled using a slight modification of the UC Davis – Rizzo Lab sampling protocol (Attachment at end), which utilizes Rhododendron leaves as "bait" for *Phytophthora* spp. Each watercourse was sampled over two sequential time periods of approximately 2-3 weeks each, with 2 replicates/watercourse site for each sampling period (a total of 4 sets of baits for each watercourse site, provided no baits were lost). Placement of baits began May 3rd and the last baits were retrieved June 30 (see Table at end of report). Most watercourses were accessed by road or trail, but some watercourses entering New Bullard's Bar Reservoir were accessed by boat. Samples were processed at UC Davis to determine if *Phytophthora ramorum* was present. Incidental to this, other *Phytophthora spp.*, were often recovered.

## **Results and Discussion**

Thirty-one watercourse sites were baited. Ultimately, a total of 118 sets of baits (see Table) were submitted for diagnostics. Watercourses were located in the following major drainages: Feather, Yuba, Bear, American, and Consumnes Rivers. *Phytophthora ramorum* was not recovered from any of the watercourses.

Spring of 2008 was exceptionally dry and marked the second consecutive dry spring. In 2007, the cumulative total inches of precipitation for the months of March, April, and May averaged for 8 northern Sierra weather stations was 5.9 inches; In 2008, it was 3.38. The historical average is 12.9 (California Department of Water Resources). It is presumed that spring precipitation would influence the recovery of *Phytophthora ramorum* from watercourses in the northern Sierra Nevada, but there is no direct evidence of this since the pathogen has never been isolated from this part of the State.

Over the past 5 years, the following surveys have been conducted in the northern Sierra Nevada:

- 2004: 23 vegetation transects in Tehama and Butte Counties
- 2005: 35 vegetation transects and 11 watercourse sites baited in Butte and Yuba Counties
- 2006: 32 vegetation transects and 23 watercourse sites baited in Butte, Yuba,

Nevada, Placer, and El Dorado Counties

- 2007: 28 watercourse sites baited in Butte, Yuba, Nevada, Placer, and El Dorado Counties
- 2008: 31 watercourse sites baited in Butte, Yuba, Nevada, Placer, and El Dorado Counties

Challenges encountered include:

- difficult terrain, access, and a huge land base
- a multitude of water impoundments throughout the survey area that serve as 'deadends' for downstream transport of *Phytophthora* propagules; hence, impoundments limit the amount of upstream habitat that is effectively surveyed by baits.
- high water levels and debris during spring runoff, especially in wet years
- dry spring weather, which potentially could limit sporulation by *Phytophthora ramorum*.

*P. ramorum* has not been detected by these surveys, however, other *Phytophthora spp.* commonly found in forest environments have been recovered from both vegetation and watercourse samples. Also, hosts for *P. ramorum* are abundant in all areas surveyed. These results support the utility of the survey methods and indicate that *P. ramorum* likely could become established in this area if it were introduced into a suitable local environment.

End of Report

## Table and Attachment follow

# Table. Location of Watercourse Sites and Sampling dates. California 2008.

	UTM	(WGS 84)					
Site Name	E	<u>`N</u>	date out	date in	date out	date in	County
Sucker Run	645854	4379459	22-May	6-Jun	6-Jun	25-Jun	Butte
Frey Creek	648452	4387332	23-May	9-Jun	9-Jun	26-Jun	Butte
French Creek	639415	4396036	24-May	10-Jun	10-Jun	30-Jun	Butte
Berry Creek Rd	634509	4391446	24-May	10-Jun	10-Jun	30-Jun	Butte
Flea Creek	632493	4407182	23-May	9-Jun	9-Jun	27-Jun	Butte
Mill Creek (Butte Co)	633770	4407182	23-May	9-Jun	9-Jun	27-Jun	Butte
Madrone Lk Berry Ck	637210	4389511	24-May	10-Jun	10-Jun	30-Jun	Butte
Galen Creek	637432	4390490	24-May	10-Jun	10-Jun	30-Jun	Butte
Bridger Creek	664071	4369303	22-May	6-Jun	6-Jun	23-Jun	Yuba
Little Oregon Cr	657085	4365909	13-May	29-May	29-May	17-Jun	Yuba
Middle Fork Yuba R	664885	4362007	13-May	29-May	29-May	6-Jun	Yuba/Nevada
Mill Creek (Yuba Co)	660977	4368406	22-May	6-Jun	6-Jun	23-Jun	Yuba
Indian Creek	657583	4371431	22-May	6-Jun	6-Jun	23-Jun	Yuba
Wolf Ck Grass Valley	670617	4343707	6-May	25-May	25-May	11-Jun	Nevada
Deer Creek	671512	4347885	6-May	25-May	25-May	11-Jun	Nevada
Steephollow Ck.	681716	4338846	27-May	14-Jun	14-Jun	24-Jun	Nevada
Willow Creek	673986	4331985	9-May	25-May	25-May	11-Jun	Nevada
South Fork Wolf Ck	669144	4330081	7-May	25-May	25-May	11-Jun	Nevada
Kentucky Creek	655343	4350553	7-May	25-May	25-May	12-Jun	Nevada
Squirrel Creek	659593	4342845	7-May	25-May	25-May	12-Jun	Nevada
Dog Bar-Bear River	672424	4325558	7-May	25-May	25-May	11-Jun	Nevada/Placer
S. Fork of the Yuba	663890	4351087	25-May	12-Jun	17-Jun	25-Jun	Nevada
Pilot Creek	702143	4312428	9-May	27-May	27-May	14-Jun	ElDorado
N Fk of Consumnes	699991	4280824	3-May	26-May	26-May	13-Jun	ElDorado
Rock Creek	693140	4296423	3-May	26-May	26-May	13-Jun	ElDorado
Weber Creek	694446	4286690	3-May	26-May	26-May	13-Jun	ElDorado
N. Fork of American	678315	4318730	12-May	28-May	28-May	15-Jun	Placer
N Fk of Mid Fk of Am	695321	4320160	4-May	28-May	28-May	16-Jun	ElDorado
Rubicon River	697107	4319271	4-May	28-May	28-May	16-Jun	ElDorado
S. Fk. Am. River	692368	4293048	26-May	13-Jun	14-Jun	21-Jun	ElDorado
M. Fk. Am. River	696396	4319846	4-May	28-May	28-May	16-Jun	Placer

#### Attachment UC DAVIS-RIZZO LAB WATERCOURSE BAIT METHODS

#### Watercourse selection:

- Sites are selected based on accessibility, local cooperation (for remote locations), minimum visibility, broadly representing county watercourses, and perennial water flow
- All watershed and watercourse sizes are considered (within reason) although accessibility during floods can be limited; we have recovered *P. ramorum* from one large river
- Each site is sampled at six week intervals year-round; adjusting sample location, time of leaves in watercourse, and replacing parts as needed
- GPS coordinates of each site are recorded to use for mapping

#### **Bait placement:**

- Make baiting bags out of approx 1mm fiberglass mesh (window screening material); cut square foot pieces and fold one edge back toward other edge, leaving approx 4" of non-overlap, and staple edges; staple five equal size pockets along the width of bag; make sure enough overlap of extra mesh to cover openings of pockets
- Clean, disease-free Rhododendron (we use Colonel Cohen horticultural variety, Gomer waterii variety also works well but any will work) leaves are placed in mesh bags
- Place bubble wrap at end slots in bags to help float bag near water surface
- Weave rope (nylon 3/16") through mesh bag to hold flap closed
- Ten leaves are placed at each location with two replicate locations per site.
- Bags are secured to riverbanks and floated near the water surface for 7-21 days with the minimum time period in warm weather and warm watercourse temperatures and longer intervals in cold conditions. Interval time adjusted year-round.
- Tie bag up high on riverbank to secure location (preferably so location is accessible during all flood stages)
- Consider attachment of 1 lb round fishing weight with highly visible and heavy gauge fishing line or use large rocks if needed to keep bag in regular watercourse flow and away from edge/bank
- Flag rope with contact info
- Clean soil/mud off boots used for accessing watercourse (rubber boots work great)- use 95% Ethanol or 10% bleach water; optional if not infested watercourse course

#### **Collection:**

- Remove leaves from water and place in separate sample collection bags
- Rinse bag and leaves in watercourse if dirt and detritus on leaves/bag/rope
- Take water temperature of watercourse at pick up- leave thermometer in water +2min (this helps evaluate how long to leave baits in watercourses)
- Sterilize removed bags in 10% bleach water for 20-30 minutes, rinse, and dry; reuse on future sampling
- Refrigerate samples prior to isolation

#### **Isolations:**

• Leaves are surface sterilized in 95% Ethanol for 30 seconds, rinsed with DI water, and air-dried for 1-2 hours. (Optional- Hansen Lab does not do this step, alternatively they just clean leaves with DI water, I like to make sure infection is on that leaf and not cross-contamination from other leaves in sample bag)

- Disease symptoms are described and recorded for all leaves.
- Symptomatic leaves are isolated onto *Phytophthora*-selective media (PARP) with 0.025g/L hymexazol, known to reduce *Pythium* species growth without impacting *Phytophthora* growth. Experiments have shown minimal inhibition of *P. ramorum* growth with this concentration of hymexazol (Fichtner *et al* 2005). Current experiments are being conducted examining hymexazol inhibition on other *Phytophthora* species. Hansen lab also uses this media.
- Submerge 10-15 leaf pieces max in media per petri-plate as flat as possible (in order to see structures forming around leaf surface) and to permit space for hyphal growth and clean transfer of organism
- Plates are incubated at 18°C

#### **Results:**

- Check plates every three to five days microscopically, carefully examining each leaf piece around entire edge for hyphae and/or reproductive structures
- Keep plates at least 3-4 weeks for late recovery of pathogens
- Any *Phytophthora*-like organisms are transferred and further examined for identification.